

Residential Customers How To Take A Sample

Sampling Instructions for Coliform and Fecal Coliform Bacteria

1. Choose a water tap which has no aerator or one which the aerator can be removed. If tap leaks it cannot be used.
2. Allow cold water to run evenly for 3 to 5 minutes.
3. Open bottle cap **DO NOT RINSE OR OVERFLOW**, bottle contains a dechlorinating agent which must be mixed with sample.
4. **DO NOT TOUCH INSIDE OF BOTTLE OR CAP.**
5. Fill sterilized container to **100ml** mark. Replace cap and press down to ensure cap is tight.
6. Return sample to laboratory within 24 hrs. of taking the sample.
Sample must be kept cold until received at laboratory.
SAMPLE must be transported on ice.

NOTE: Payment is due when samples are received at laboratory.
Acceptable payment methods Cash, Check, Credit Card

Sample will be refused if less than 100ml.
Bacteria samples are accepted Monday – Thursday 8a.m. to 4p.m.
No Bacteria samples are accepted on Friday

Sampling Instructions for Drinking Water VOC's

NOTE: 1:1 HCL and a pasture pipet will be needed for this sampling.
Laboratory provides 3 sample vials that contain Ascorbic Acid and one vial that is a Trip Blank. All four vials should be returned to the Laboratory.

1. Allow cold water to run for approximately 5 minutes
2. Reduce flow to a slow steady stream.
3. Fill vial half way, add 2 drops of 1:1 Hydrochloric acid, and continue filling vial to extreme top. **DO NOT OVERFLOW.**
4. Replace cap and tighten.
5. Follow steps 3 & 4 to fill other two vials.

6. Flip sample bottles upside down to ensure that there are **NO** air bubbles. If air bubbles are present sample cannot be accepted at lab.
7. Keep samples at 4°C from time of sampling until delivered to laboratory.
8. Samples must be received at lab within 24hrs.

Sampling Instructions for Copper and Lead

NOTE:

- After selecting location that sample will be taken from, allow tap to sit unused for a minimum of 6 hours for most accurate results.
- It is recommended that sample be taken as the first draw of the day or upon returning home from work. This would allow for the water to be unused for a minimum of 6hrs.
- If possible a kitchen or bathroom tap would be best sampling point.
- Sample will not be accurate if sample is taken from a tap that is hooked up to a water softener. If kitchen tap is hooked up to a water softener than take sample from bathroom tap or another tap that is not hooked up to the water softener.

1. Do not remove aerator prior to sampling.
2. Place open bottle under tap and slowly open the cold water tap. Fill sample bottle to 1000ml line. (If no line is provided fill to top).
3. Turn water off and replace cap and tighten to ensure sample does not spill.
4. Label bottle and complete chain of custody with all required information.
5. Please include any plumbing work recently done or if sample had to be taken from a tap that is hooked up to a water softener.
6. Keep sample cold from time of sampling until delivery to laboratory.
7. Sample must be received at lab within 24hrs.

Sampling Instructions for Nitrates

1. Allow water to run for approximately 1 minute.
2. Fill 8oz. bottle to top replace cap and tighten.
3. Keep sample cold from time of sampling until delivery to laboratory.
4. Sample must be received at lab within 24hrs.

Sampling instructions for Iron, Manganese, Corrosivity and FHA/VHA Loan

NOTE: Aerator does not need to be removed.

Allow cold water to run for approximately one minute.

1. Fill sample bottle to 1000ml mark (if no mark is provided fill bottle to top).
2. Replace cap and tighten.
3. Keep sample cold until delivered to laboratory.
4. Sample must be received at lab within 24hrs.

Sampling instructions for TTHMs

NOTE: 1:1 HCL and a pasture pipet will be needed for this sampling.

Laboratory provides 3 sample vials that contain Ascorbic Acid and one vial that is a Trip Blank. All four vials should be returned to the Laboratory.

1. Allow cold water to run for approximately 5 minutes
2. Reduce flow to a slow steady stream.
3. Fill vial half way, add 2 drops of 1:1 Hydrochloric acid, and continue filling vial to extreme top. **DO NOT OVERFLOW.**
4. Replace cap and tighten.
5. Follow steps 3 & 4 to fill other two vials.
6. Flip sample bottles upside down to ensure that there are **NO** air bubbles. If air bubbles are present sample cannot be accepted at lab.
7. Keep samples at 4°C from time of sampling until delivered to laboratory.
8. Samples must be received at lab within 24hrs.

Sampling instructions for HAA5's

Note: Aerator does not need to be removed.

1. Allow cold water to run for approximately 5 minutes.
2. Reduce flow of water to a steady stream.
3. Fill 1 250ml Amber jar containing 0.025 g. of Ammonium Chloride.
4. Replace lid, and make sure it is secure.
5. Keep samples at 4°C from the time of sampling until received at the laboratory.
6. Samples must be received at lab within 24hrs.